**Required** **Formatting**

* *File Type:* The manuscript must be submitted as an editable .doc or .docx file.
* *Font:* 12 pt, Calibri.
* *Line spacing:* Include single-line spaces between all paragraphs, headings, steps, *etc.*
* *Page margins:* 1 inch (2.54 cm) on all sides.
* *Page size:* Standard US Letter.

***Technical Language***

* The manuscript text must be **original, in complete sentences, and in paragraph form**.
* Define acronyms/abbreviations upon first use in the main text.
* Use SI abbreviations for all units: L, mL, µL, h, min, s, *etc.*
* Italicize all Latin words and nomenclature: *et al., in vivo, in vitro, in silico, i.e., e.g., etc.*
* Include a space between all numbers and the corresponding unit: 50 mg, 100 mL, 37 °C, *etc.*
* List all centrifugation speeds in terms of centrifugal g-force instead of rpm: 100 x g
* Molecular formulas should include subscripts: CO2, H2O2, O2, *etc.*
* Abbreviate species names after first use: *Caenorhabditis elegans* should be *C. elegans.*

**TITLE:** *(150 characters maximum)*

**The title should represent the content included in the video** and include the model system used or the type of study design. Please avoid the use of abbreviations.

An open-source EEGLAB plugin for jointly analyzing EEG and cardiovascular signals.

**AUTHORS AND AFFILIATIONS:**

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**KEYWORDS:** *(6 minimum, 12 maximum)*

EEGLAB, EEG, ECG, PPG, HRV, joint analysis, feature-based, event-related

**SUMMARY:** *(10-word minimum, 50-word maximum)*

The summary should clearly state the goal of the protocol. It may include a general description of the method and its applications. This description should focus on the protocol, not the results obtained by the method.

Enter text here.

**ABSTRACT:** *(150-word minimum, 300-word maximum)*

The abstract should focus on the method being presented rather than the results of a specific experiment. Include a statement about the purpose of the method. A more detailed overview of the method and a summary of its advantages, limitations, and applications is appropriate. Please focus on the general types of results acquired. Do not include references here.

We present an open-source toolbox for processing and analyzing joint electroencephalography (EEG) and electrocardiography/photoplethysmography (ECG/PPG) signals. EEG and ECG/PPG are non-invasive, low-cost, and highly mobile techniques for monitoring brain and cardiovascular activity. They offer distinct advantages over other neuroimaging methods, such as magnetoencephalography (MEG), functional magnetic resonance imaging (fMRI), and positron emission tomography (PET) in terms of their lower cost, portability, and ability to collect data in real-world settings. The interaction between the cortical and cardiovascular systems has been the subject of increasing interest, as it can capture useful information about the coupling of brain and cardiovascular function. However, analyzing the interaction between these time series can be challenging due to the complex or noisy nature of the data and the lack of standardized analysis methods. Existing tools for jointly analyzing these biosignals often require a high level of technical expertise and may not provide all of the necessary features for studying the interaction between these signals. Moreover, many existing tools are not open-source, limiting their accessibility, and a consensus towards clear signal processing guidelines is still in development and reproducibility. The proposed toolbox addresses these limitations by providing a comprehensive set of signal processing functions, implementing the latest guidelines as its default parameters, and three main functions: 1) feature-based for continuous data for assessing associations between various features extracted from EEG and HRV signals (time, frequency, and nonlinear features); 2) event-related for epoched data (EEG-ECG coherence, heartbeat-evoked potentials in time, frequency, and time-frequency domains); 3) EEG signal processing (extracts heart components from EEG signals using ICA). Robust statistics are implemented for assessing relationships between these time series at the individual level (e.g., permutation statistics, corrections for type 1 error). Linear and nonlinear features are provided to better capture complex, bidirectional interactions between that take place between the cardiovascular, subcortical, and cortical systems. Additionally, the toolbox can be used simply for extracting heart components from the EEG data using independent component analysis (ICA). This open-source toolbox should provide a valuable resource for researchers and clinicians interested in studying the relationship between the brain and cardiovascular activity and can be easily adapted and extended to meet specific research needs.

**INTRODUCTION:** *(150-word minimum, 1500-word maximum, 2-paragraph minimum)*

This section should include:

* A clear statement of the purpose of this method
* The rationale behind the development and/or use of this method
* The advantages over alternative methods with references to relevant studies
* The context of the method in the wider body of published literature
* Information to help readers decide whether the method described is appropriate for them
* Purpose of this method

The purpose of this method is to ease the investigation of relationships between the brain and the heart by facilitating the joint analysis of electroencephalography (EEG) and cardiovascular signals, namely electrocardiography (ECG) and photoplethysmography (PPG), while implanting the latest guidelines from experts in this field. This tool addresses limitations from existing tools and is made open source to facilitate accessibility and reproducibility in the field. The proposed toolbox should serve as a valuable resource for researchers and clinicians interested in removing cardiac artifacts from EEG signals, in extracting features from EEG and ECG/PPG signals, or in studying the relationship between brain and cardiovascular activity. Ultimately, this toolbox aims to pave the way for more in-depth investigations into the complex interplay between the brain and heart systems.

* The rationale behind the development and/or use of this method + context of the method in the wider body of published literature

**Reductionist vs. multimodal approaches**

For a long time, the reductionist approach has dominated scientific inquiry in the field of human physiology and cognition. This approach involved dissecting complex bodily and mental processes into smaller, more manageable components, allowing researchers to focus on individual systems in isolation. This strategy arose due to the immense challenge of studying the intricate and interconnected nature of the human body and mind (von Bertalanffy, 1968). Reductionism has been instrumental in understanding individual subsystems in isolation, such as elucidating the role of ion channels and action potentials for neural (Hodgkin & Huxley, 1952) and cardiac (Bean et al., 1984) communication. However, a large gap remains in our understanding of how these isolated systems interact together on a larger spatial and temporal scale. Thanks to recent advances in technology, a more integrative approach is now gaining interest. Innovations in hardware, signal processing, data storage, and computing capabilities have enabled researchers to collect and analyze electrophysiological signals from different bodily systems simultaneously (e.g., heart, brain, skin, etc.; Jiang et al. 2019; Insel et al., 2017; Kappenman & Luck, 2010). This integrative – or multimodal – approach will pave the way for a more comprehensive understanding of human physiology, cognition, pathology, and consciousness (Bunge, 2003; Fuchs, 2018; Thayer & Lane, 2009). Consequently, this multimodal approach is now considered an essential component of modern research, complementing traditional reductionist methods and offering novel insights into the synergistic mechanisms governing the human body and mind.

**Heart-brain research with fMRI and PET**

Studying the intricate relationship between the brain and the heart can yield valuable insights into the underlying physiology and anatomy of the human body, ultimately leading to the development of novel diagnostic and therapeutic tools. The relationship between the heart and the brain has been studied via neuroimaging methods such as functional magnetic resonance imaging (fMRI) and positron emission tomography (PET). Using these tools, researchers highlighted some brain regions associated with cardiovascular control (e.g., manipulation of heart rate and blood pressure; Critchley 2000; Napadow 2008), showed the influence of heart rate on the BOLD signal (Chang et al., 2009; Shmueli 2007), or identified potential brain-body pathways contributing to coronary heart disease (i.e., stress-evoked blood pressure; Gianaros 2009).

**EEG, ECG, and PPG**

While these studies have significantly advanced our understanding of the complex interplay between the central nervous system (CNS) and cardiovascular function, these neuroimaging techniques are expensive, have limited availability, and are confined to controlled laboratory settings, which restricts their practicality for real-world and large-scale applications. In contrast, EEG and ECG/PPG are more affordable and portable tools that offer the potential for studying brain-heart interactions in more diverse settings and populations or over longer periods of time, providing new opportunities for investigating the dynamic relationship between brain and heart function. ECG measures the electrical signals generated by the heart when it contracts and relaxes via the placement of electrodes on the skin (usually on the chest, arms, or legs). PPG measures blood volume changes in the microvascular tissues by using a light source (e.g., LED) and a photodetector placed on the skin (commonly on a fingertip, earlobe, or forehead). Since blood absorbs more light than the surrounding tissue, the PPG signal can be used to estimate blood flow and pulse rate. Both methods provide valuable information about the cardiovascular function, but they serve different purposes and offer distinct types of data. As such, the use of EEG and ECG/PPG holds great promise for advancing our understanding of the physiological, cognitive, and emotional processes underlying brain-heart interactions and their implications for human health and well-being. Similar to ECG, EEG records the electrical fields generated by synchronized activity of thousands of cortical neurons by placing electrodes on the scalp.

**The 2 approaches to jointly analyzing EEG and cardiovascular signals**

There are two main approaches to study interactions between EEG and cardiovascular signals:

1. Event-related: each heartbeat is marked in the EEG time series to examine with high temporal accuracy the cortical activity processing cardiac signals (Schandry 1981; Pollatos 2005; Montoya 1993; Park & Tallon-Baudry 2014; Couto 2015; Jiang et al. 2019; Dirlich et al., 1998). This method is termed heartbeat-evoked potentials (HEP) and is similar to traditional event-related potential (ERP) studies, which required the two time series to be time-locked, but the events are heartbeats as opposed to stimuli.
2. Feature-based: using continuous data, this approach extracts features from the EEG signals and from the cardiovascular signals and examines associations between them. Cardiovascular features are typically heart-rate variability (HRV) measures in the time, frequency, or nonlinear domains. This has been done with ECG (Thayer et al. 2012; Mather 2018; Kemp 2013) and PPG to a lesser extent (Khosrow-Khavar 2014). This approach provides trait information that can be used for making medical forecasting or classification (e.g., mental or physical health) and finding more general associations relative to HEP that focus on mechanisms at millisecond accuracy.

The advantages over alternative methods with references to relevant studies

While some tools exist to process cardiovascular and EEG (e.g., EEGLAB, Fieldtrip, Brainstorm) signals separately, none is currently available for jointly analyzing them. Furthermore, the tools available to process cardiovascular signals:

* require expensive license purchase and do not allow processing large datasets in batch via command line (Kubios);
* or require advanced programming skills by not providing a graphical user interface (GUI; Physionet Cardiovascular Signal toolbox; [Vest et al. 2019](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6442742/pdf/nihms-1509712.pdf)).

To our knowledge, three open-source MATLAB toolboxes exist to support HEP analysis with a GUI. The ecg-kit ([REF](https://github.com/marianux/ecg-kit); Demski and Soria, 2016), the HEPLAB EEGLAB plugin ([REF](https://github.com/perakakis/HEPLAB)), and the CARE-rCortex EEGLAB plugin ([Grosselin et al. 2018](https://www.sciencedirect.com/science/article/abs/pii/S0165027018302474)). While HEPLAB and ecg-kit facilitate HEP analysis by detecting heartbeats and marking them in the EEG signals, they do not provide statistical tools for analysis, and are limited to the time domain (i.e., ERP). The CARE-rCortex addressed these issues by supporting both ECG and respiratory signals, time-frequency domain analysis, statistics, and advanced baseline normalization and correction methods specifically adapted to HEP analysis. However, it uses the Bonferoni method for multiple comparisons correction, which is too conservative for EEG analysis (Ref), and it does not offer command-line access for processing and analyzing large datasets. Furthermore, recent studies recommend against baseline correction methods, highlighting how they reduce signal-to-noise ratio (SNR) and are “statistically unnecessary and undesirable” ([Alday 2019](https://onlinelibrary.wiley.com/doi/abs/10.1111/psyp.13451); Delorme 2023).

To address these limitations, this article presents a new open-source MATLAB toolbox, named “BrainBeats”, implemented as an EEGLAB plugin designed to jointly process and analyze EEG and ECG/PPG signals. It incorporates the following advantages over previous methods:

* Easy-to-use GUI (for non-programmers) and command line mode (for programmers aiming to perform automated processing and analysis on large datasets, use more advanced parameters).
* Implements the validated algorithms from the Physionet Cardiovascular Signal toolbox ([Vest et al. 2019](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6442742/pdf/nihms-1509712.pdf)) for detecting the QRS complexes, estimating a validated signal quality index (SQI) of the RR time series, and removing or interpolating the RR artifacts automatically with various algorithms (e.g., linear, cubic, nearest neighbor, etc.) to otbain the normal-to-normal (NN) intervals.
* Implements validated EEG signal processing, including referencing (to infinity for montages with at least 30; Yao [2001](https://pubmed.ncbi.nlm.nih.gov/11761077/), [200](https://pubmed.ncbi.nlm.nih.gov/15798293/)5; [2017](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5331115/); [Dong 2017](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5670162/)), detection and interpolation of bad channels, detection and interpolation of EEG artifacts (Mullen 2015; Delorme 2004; 2023; Pion-Tonachini 2019). Note: Users can also use the toolbox with already cleaned data.
* Can be used to automatically extracting heart components from EEG signals using the ICLabel EEGLAB plugin (**Method 1**).
* Supports HEP analysis (**Method 2**) in time, frequency, and time-frequency domains (EEGLAB STUDY mode) and advanced statistics using hierarchical linear modeling provided by the LIMO plugin (Pernet 2011). This statistical approach accounts well for within and between- subjects variance, downweighs artifactual trials using weighted least square (WLS; Pernet 2022) optimization, and applies bootstrap statistics and advanced corrections for multiple comparisons (e.g., spatiotemporal cluster correction). Furthermore, inter-trial coherence analysis can be performed using EEGLAB default statistics.
* Supports for the first time the study of brain-heart associations using a feature-based approach (**Method 3**). HRV features (listed in **Figure 1**) are calculated following guidelines to facilitate standardized and accurate analysis (Shaffer and Ginsberg 2017). The novelty about this approach is that it captures the trait variable and can be used on large continuous EEG datasets collected in the laboratory or in real-world settings using wearable headsets for example (e.g., clinics, patient’s home; Cannard et al., 2020). These features can be analyzed with simple correlation procedures or with machine learning to make classifications or predictions (e.g., detection of depression before symptoms are too advanced; REF).
* Provides various data visualizations to inspect signals (raw, RR intervals, EEG and RR artifacts that have been removed or interpolated) or outputs for each file (e.g., HEP data, power spectra from NN or EEG signals, etc.; see **Figures 2 and 3**).

Information to help readers decide whether the method described is appropriate for them

This toolbox is appropriate for any researcher or clinician that possesses data containing EEG and ECG/PPG signals. The plugin does not support the importation of EEG and ECG/PPG signals from separate files yet (although this feature will be available soon). The toolbox can be used for simply removing heart artifacts from EEG signals with high confidence, to simply extract EEG or HRV features (for other purposes), or for assessing interactions between these two systems.

**PROTOCOL:** *(1-page minimum, 10-page maximum)*

**The protocol text should provide a detailed description to enable the accurate replication of the presented method (including setup, materials, actions, conditions, *etc.*) by both experts and researchers new to the field.** Well-established methods (*e.g.*, Western Blotting, PCR, *etc.*) used within the protocol should be cited as necessary and any modification of the aforementioned procedures should be described.

**Format:**

* The protocol must be a numbered list: step 1 followed by 1.1, followed by 1.1.1, *etc*.
* Include a space between each numbered step or note in the protocol.
* Each step should include 1-2 actions and contain 2-3 sentences. Use sub-steps as necessary.
* Please do not use indentations.
* Do not underline any text in the protocol; however, **bold** text is acceptable for emphasis.

**Grammar:**

* Use complete sentences throughout the protocol.
* Avoid the use of personal pronouns or colloquial phrases (*e.g.*, I, you, your, we, our).
* Use the active/imperative voice throughout this section.

Good Example: Add 30 µL of solution A to 30 µL of solution B.

Bad Example: 30 µL of solution A was added to 30 µL of solution B.

* Avoid the use of commercial language, including ™/®/© symbols or company brand names before/after an instrument or reagent. Cite these in the Table of Materials instead.

**Technical Specifications:**

* Use subheadings for clarity if there are discrete stages in the protocol.
* Please indicate any points at which the experiment can be paused and then restarted later. For these situations, indicate the choices at that point in the protocol.

Example: Incubate the filter for 4 h at room temperature or overnight at 4 °C.

* Indicate any toxic or harmful chemicals with the word “CAUTION” when they are first used, and include notes that describe the hazard and the appropriate handling guidelines.
* All methods that involve the use of human or animal subjects and/or tissue or field sampling must include an ethics statement before the numbered protocol section (see Editorial Policies: ([www.jove.com/author/editorial-policies](http://www.jove.com/author/editorial-policies)) for more information).

Example: All methods described here have been approved by the Institutional Animal Care and Use Committee (IACUC) of Harvard University.

**Protocol Length**:

There is a 10-page limit (with proper formatting) for the amount of text written in the protocol section. There is a 2.75-page limit on the amount of content we can film for a single video article. **For a Protocol section that exceeds 3 pages, highlight in yellow up to 2.75 pages (no less than 1 page) of protocol text (including headers and spacing) to be featured in the video. Our scriptwriters will derive the video script directly from the highlighted text.**

* + - Bear in mind the goal of the protocol, and highlight the critical steps to be filmed.
    - Please ensure that the highlighted steps form a cohesive narrative with a logical flow from one highlighted step to the next. The highlighted part of the step must include at least one action that is written in the imperative voice.
    - You do not need to delete steps from the protocol. The full-length manuscript will be published along with the video. All un-filmed steps will still be available in the written manuscript for readers.

**Equations**:(Example of JoVE video with equations: [www.jove.com/51288](http://www.jove.com/51288))

* Ensure all inline equations are formatted identically, using the consistent font style.
* Separate each equation to its own line, and define all terms in the equation. A 4-line equation should take up 4x the vertical space as a single line equation.
* Do not embed equations as images. Instead, use the equation builder in MS Word.

Enter text here (10-page maximum, 2.75 pages of highlighted text for filming).

**METHOD 1: Remove heart components from EEG signals**

1.1. Load the 1st sample dataset containing both 3 EEG channels and one ECG channel into EEGLAB: File > Load existing dataset > select “sample\_data1.set” > Open. Note: You may need to install other EEGLAB plugins to import your dataset depending on the file format (e.g., .bdf, .edf, .vhdr, etc.).

1.2. Select parameters: Tools > Run BrainBeats (see **Figure 1.1.**). The general user interface (GUI) pops up (**Figure 1.2.**). Select analysis type as “Remove heart components from EEG signals”, “ECG” heart data type, the name of the ECG channel “ECG” from the list of channels, and “No (already processed)” as this sample file was already preprocessed. Note: The “Plot outputs” option is set by default. Click “Ok” to launch.

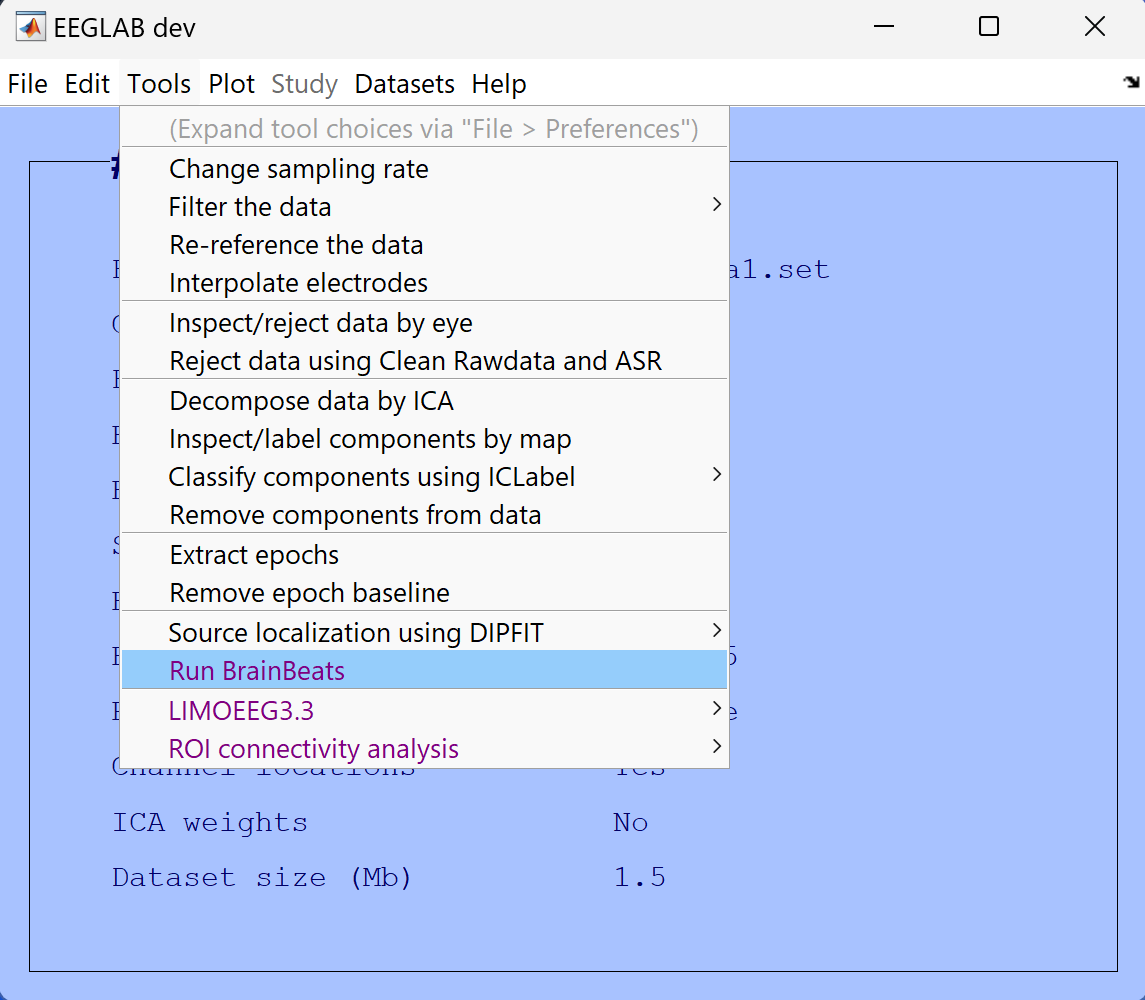


Figure 1.1. Launching the BrainBeats plugin from the mean EEGLAB menu.

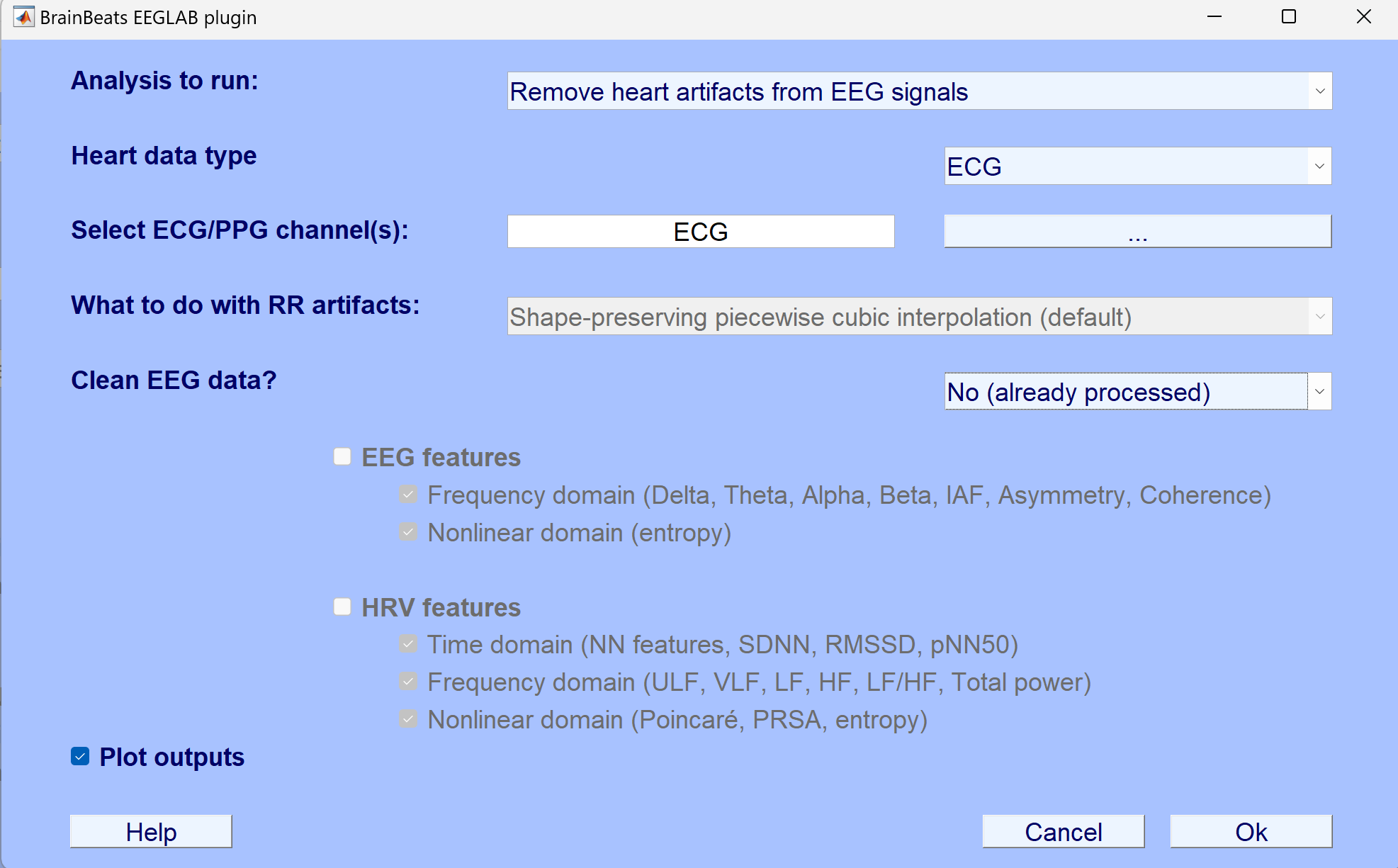
****

Figure 1.2. Selecting parameters from the general user interface (GUI).

1.3. Independent component analysis (ICA) runs (Infomax algorithm), using PCA reduction is the effective data rank is lower than the number of channels ([Ref](https://www.frontiersin.org/articles/10.3389/frsip.2023.1064138/full?&utm_source=Email_to_authors_&utm_medium=Email&utm_content=T1_11.5e1_author&utm_campaign=Email_publication&field=&journalName=Frontiers_in_Signal_Processing&id=1064138)), and the ICLabel plugin (Ref) is called to automatically detect heart components with at least 95% confidence. Note: the ECG channel(s) is kept, increasing ICA’s source separation performance and chances to separate heart components from the EEG signals, if present. See representative results below.

These steps can be run automatically over many files using the following command lines:

eeglab; close;

dataDir = fileparts(which('pop\_BrainBeats.m'));

EEG = pop\_loadset('filename','sample\_data1.set','filepath',dataDir);

EEG = pop\_BrainBeats(EEG,'analysis','rm\_heart','heart\_signal','ECG','heart\_channels', {'ECG'},'vis',true);

**METHOD 2: Heartbeat-evoked potentials (HEP)**

2.1. Load data into EEGLAB: File > Load existing dataset > select “sample\_data2.set

2.2. Tools > Run BrainBeats to select the following parameters: Click on “Heartbeat-evoked potentials HEP)” analysis to unlock the other parameters below; select the “EXG5” and “EXG6” ECG channels (click the button to display the list of channels, or type the channel names manually in the text box); You may leave the other default parameters. See overview in **Figure 2.1**.

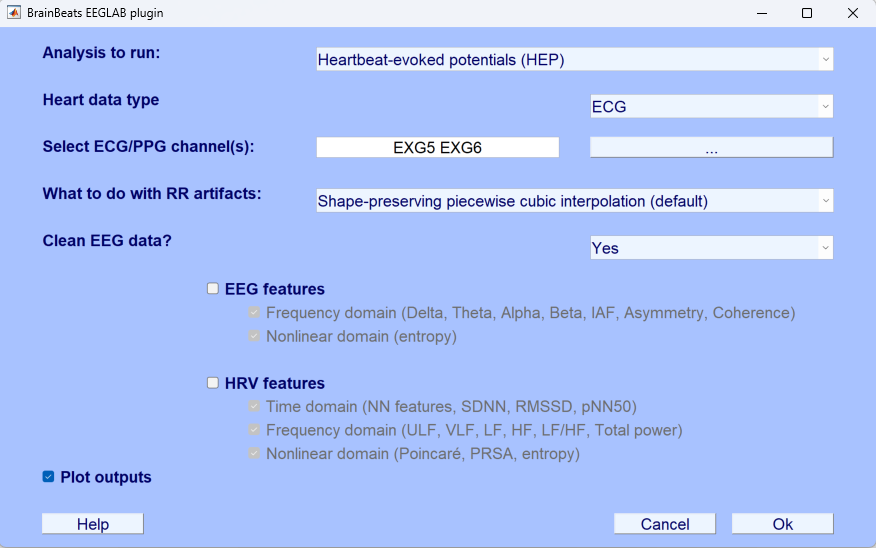


Figure 2.1. Selecting parameters from the GUI for performing heartbeat-evoked potentials (HEP) analysis.

2.3. By default, BrainBeats applies a zero-phase non-causal FIR filter to remove low-frequency drifts and high-frequency noise (high-pass cutoff frequency = 0.5 Hz, low-pass cutoff = 45; order = 846; transition bandwidth = 1 Hz). Then, BrainBeats uses the *clean\_rawdata* plugin to remove bad EEG channels (ignoring the ECG channels) and interpolates them using EEGLAB’s spherical splines interpolation (Perrin et al., 1989). When data visualization is ON, the removed channels are plotted (**Figure 2.2.**, in red).

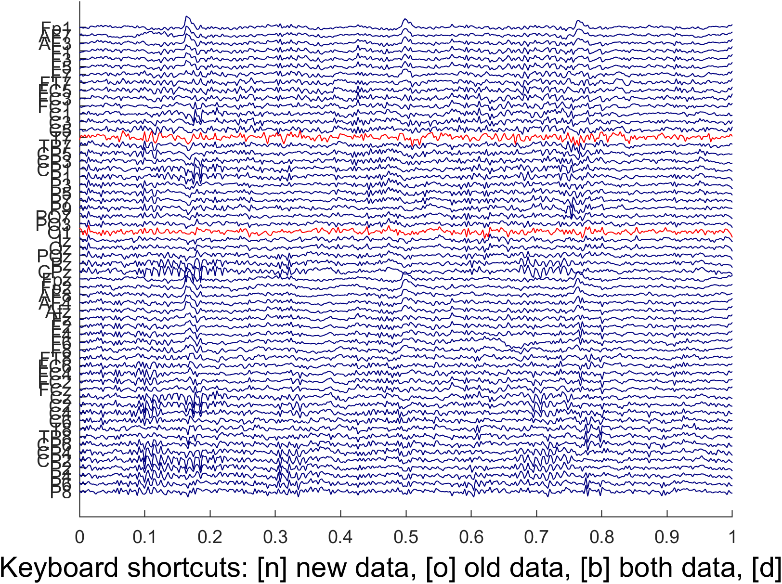
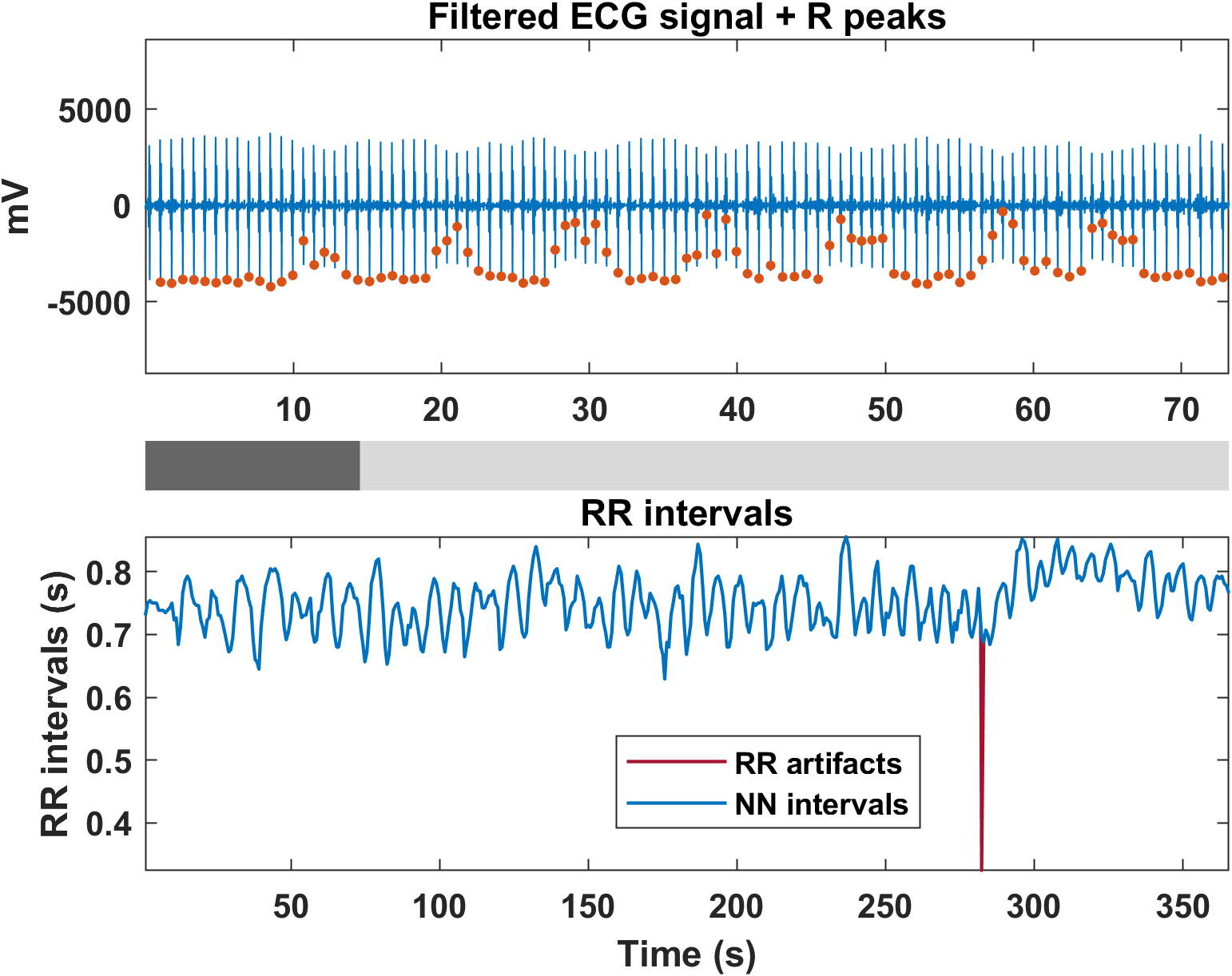


Figure 2.2. Abnormal EEG channels automatically detected and removed by EEGLAB.

2.4. The QRS complexes are detected automatically using the Pan–Tompkins method ([ref](https://ieeexplore.ieee.org/document/4122029)). The energy threshold is estimated at 98-99% of the amplitude of the distribution to better deal with large bumps. A bandpass filter is applied to the ECG signal for best performance. A search back algorithm finds missed peaks by lowering the threshold in an area where the RR interval variability is higher than 1.5 times the median. The algorithm also detects the polarity of the peaks (positive or negative). Refs: Behar et al (2014); Johnson et al. (Year). Users can scroll through the RR intervals to inspect the detected QRS complexes, and the RR artifacts that were interpolated (in red) to obtain the NN intervals (**Figure 2.3.**).

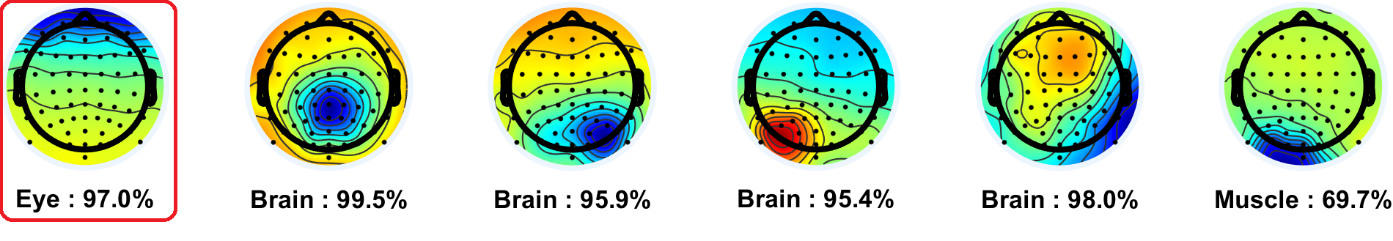
2.5. The signal quality index is calculated using the method developed by Vest et al. in the Physionet Cardiovascular Signal toolbox (Vest et al 2017). When several ECG channels are present, these steps are performed on all of them, and the channel with the best SQI is selected for the following steps.

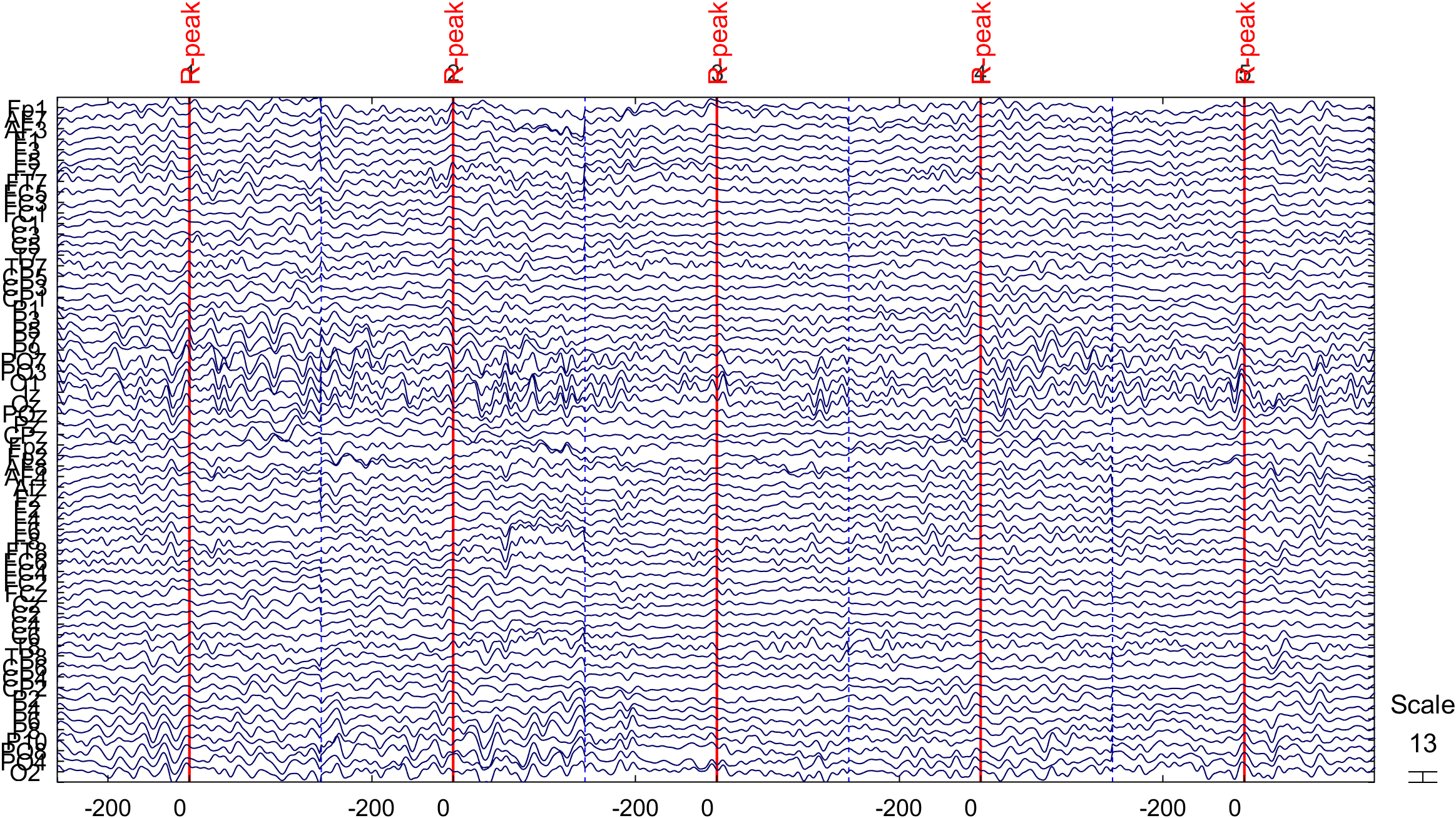
2.6. RR artifacts are interpolated to obtain NN intervals (default method is the shape-preserving piecewise cubic interpolation).



**Figure 2.3**. Top: filtered ECG time series (blue) and the detected R-peaks (orange dots). A scrolling bar allows users to go through the file in more detail. Bottom: Corresponding NN intervals (blue) after the RR artifacts were interpolated by the algorithm (red).

2.7. BrainBeats then marks the R peaks as events in the EEG signals and segments the data using the smallest interval between two R peaks in the series. BrainBeats the root mean square amplitude and a signal-to-noise ratio (SNR) measure for each epoch, and removes bad trials using the *isoutlier* MATLAB function (‘mean’ method for amplitude, and ‘grubbs’ method for SNR). BrainBeats then runs infomax ICA (with PCA dimension-reduction when the data are rank-deficient; Kim et al. 2023) and ICLabel to remove ocular and muscular components with at least 95% confidence (see **Figure 2.4. Top panel**). The final EEG time series, cleaned and epoched around the heartbeats is displayed when “Plot outputs” is set to ON (see **figure 2.4. bottom panel**).





**Figure 2.4. Top**: Ocular component classified by ICLabel. **Bottom**: EEG data segmented around the R peaks after removing bad trials, ocular components with at least 95% confidence.

2.8. Files are then saved in the same directory and with the same name as the original file with “\_HEP” at the end. Note: it is recommended to use a folder for each subject for better organization.

2.9. Users can pause before processing the next file. When all files are processed .and are later imported into an EEGLAB STUDY to compute time-frequency decompositions.

2.10. Perform hierarchical linear modeling using the LIMO plugin. A full tutorial is available at <https://github.com/LIMO-EEG-Toolbox/limo_tools/wiki>

Perform all the above steps with the following command line:

eeglab; close;

dataDir = fileparts(which('pop\_BrainBeats.m'));

EEG = pop\_loadset('filename','sample\_data2.set','filepath', fullfile(dataDir,'sample\_data'));

pop\_BrainBeats(EEG,'analysis','hep','heart\_signal',{'ECG'},'heart\_channels',{'EXG5' 'EXG6'},'clean\_eeg',true,'vis',true);

**METHOD 3: Feature-based analysis from continuous data**

3.1. Load the same dataset as in METHOD 2 into EEGLAB.

3.2. Select parameters from the GUI (Figure : Go to Tools > Run BrainBeats > Select “Extract features

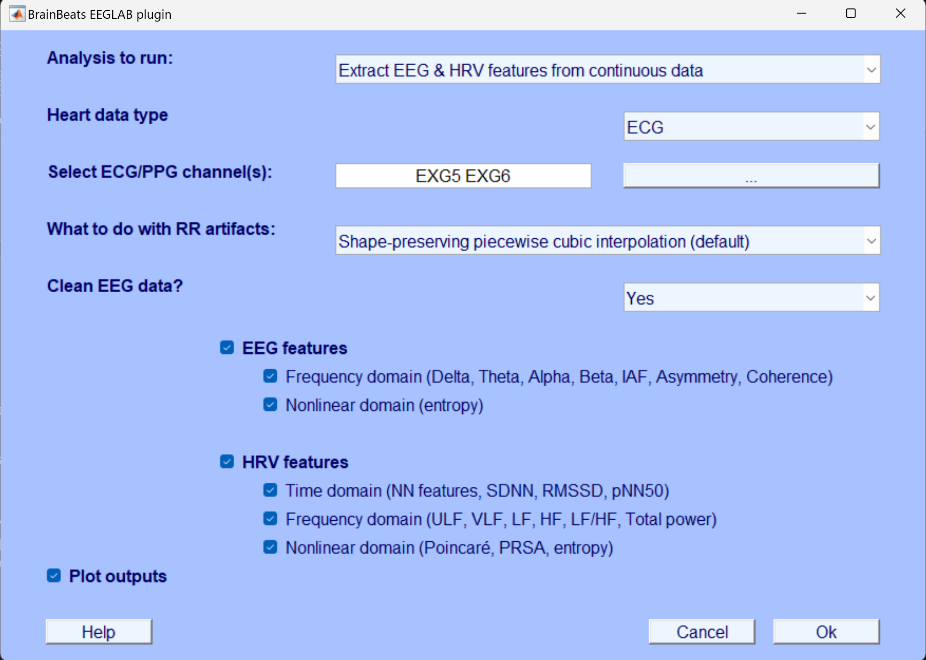


Figure 3.1. GUI to select parameters for extracting HRV and EEG features from continuous data.

Tools > Run BrainBeats to select the following parameters: Click on “Heartbeat-evoked potentials HEP)” analysis to unlock the other parameters below; select the “EXG5” and “EXG6” ECG channels (click the button to display the list of channels, or type the channel names manually in the text box); You may leave the other default parameters. See overview in **Figure 3**.

**REPRESENTATIVE RESULTS:** *(Example Representative Results section:* [*www.jove.com/52010*](http://www.jove.com/52010)*)*

Please provide a concise, written description of a representative outcome following the use of this protocol, so that a viewer will have a sense of a “positive” and/or “negative” result. **Please reference all data and figures in the manuscript**, emphasizing how the results confirm the success of the protocol, and how to interpret the data. Please include data from successful experiments, and data from sub-optimal experiments to demonstrate the range of outcomes possible. Also include results for possible outcomes if critical steps are not followed. A diagram/schematic of the method is recommended but is NOT sufficient.

All claims of the effectiveness of a method must be supported with data, *i.e.*, representative results. For example: If authors claim that method X cleanly purifies nuclear envelope proteins from a cell, they must include a figure definitively demonstrating this purification. The manuscript must include at least one figure or table providing Representative Results.

Provide a separate file for each figure and table; do NOT embed figures or tables within the manuscript document. The default placement for all figures and results tables in the final publication is below the Representative Results text. Please indicate, via brackets [Place Figure 1 here], if you prefer figure/table placement at another location in the text.

**If a figure is adapted or republished from a previous publication, authors must cite the original article in the figure legend. Reprint permission for the previously published/adapted figure is required upon acceptance.**

Example: This figure has been modified from [citation].1

**METHOD 1**

When no heart components are detected, users are informed in MATLAB’s command window, and the program ends. If at least one heart component is detected, its scalp topography and confidence level are displayed in a figure (**Figure 3 Left**). Click on the red button to visualize the heart component’s detailed properties (**Figure 3 Middle**). Click “Ok” or exit buttons when done with visualization. The ECG channel is then removed from the dataset and the heart component is extracted from the EEG signals automatically. The difference between before and after heart component removal is displayed automatically (**Figure 3 Right**). Note: All figures are only created when the “Plot outputs” box is checked in the GUI).

Graphical user interface, application

Description automatically generated Graphical user interface, application

Description automatically generated A picture containing graphical user interface

Description automatically generated

Figure 3. Left: Scalp topography and confidence level of the heart component that was detected by ICLabel. Middle: Detailed properties of the component can be visualized by clicking on the red button on the Left. Right: Difference between before and after removal of the heart component from EEG signals. The ECG channel is then removed from the dataset.

**FIGURE AND TABLE LEGENDS:** Text: Use Calibri, 20 pt. font or greater.

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Entropy features are particularly promising for capturing complex, bidirectional interactions between cardiovascular, subcortical, and cortical systems that may be hidden in nonlinear feedback loop dynamics (Costa et al., 2005).

**ACKNOWLEDGMENTS:** Please list acknowledgments and all funding sources for the work here. Also consider listing any person appearing in the film who does not appear in the authors list.

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Kioh, L.G. *et al.* *Physical Treatment in Psychiatry*. Blackwell Scientific Pubs. Boston (1988).

von Bertalanffy, L. (1968). General system theory: Foundations, development, applications. George Braziller.

Bunge, M. (2003). Emergence and convergence: Qualitative novelty and the unity of knowledge. University of Toronto Press.

Fuchs, T. (2018). Ecology of the brain: The phenomenology and biology of the embodied mind. Oxford University Press.

Thayer, J. F., & Lane, R. D. (2009). Claude Bernard and the heart-brain connection: Further elaboration of a model of neurovisceral integration. Neuroscience and Biobehavioral Reviews, 33(2), 81-88.

Insel, T. R., Landis, S. C., & Collins, F. S. (2017). Research priorities. The NIH BRAIN Initiative. Science, 340(6133), 687-688.

Kappenman, E. S., & Luck, S. J. (Eds.). (2010). The Oxford handbook of event-related potential components. Oxford University Press.

Costa, M., Goldberger, A. L., & Peng, C. K. (2005). Multiscale entropy analysis of complex physiologic time series. Physical Review Letters, 89(6), 068102.

Kemp, A. H., Quintana, D. S., Gray, M. A., Felmingham, K. L., Brown, K., & Gatt, J. M. (2010). Impact of depression and antidepressant treatment on heart rate variability: A review and meta-analysis. Biological Psychiatry, 67(11), 1067-1074.

Lehrer, P., Gevirtz, R., & Eddie, D. (2021). Heart rate variability biofeedback: A theoretical perspective and review of its effectiveness for

Bean, B. P., Cohen, C. J., & Tsien, R. W. (1984). Lidocaine block of cardiac sodium channels. The Journal of General Physiology, 73(6), 679-696.

Thayer, J. F., Åhs, F., Fredrikson, M., Sollers III, J. J., & Wager, T. D. (2012). A meta-analysis of heart rate variability and neuroimaging studies: Implications for heart rate variability as a marker of stress and health. Neuroscience & Biobehavioral Reviews, 36(2), 747-756.

Mather, M., & Thayer, J. F. (2018). How heart rate variability affects emotion regulation brain networks. Current Opinion in Behavioral Sciences, 19, 98-104.

Kemp, A. H., & Quintana, D. S. (2013). The relationship between mental and physical health: Insights from the study of heart rate variability. International Journal of Psychophysiology, 89(3), 288-296.

Jiang, H., He, B., Guo, X., Wang, X., Guo, M., Wang, Z., ... & Cui, D. (2020). Brain–Heart interactions underlying traditional Tibetan Buddhist meditation. *Cerebral Cortex*, *30*(2), 439-450.

Schandry, R. (1981). Heart beat perception and emotional experience. Psychophysiology, 18(4), 483-488.

Pollatos, O., Kirsch, W., & Schandry, R. (2005). Brain structures involved in interoceptive awareness and cardioafferent signal processing: a dipole source localization study. Human Brain Mapping, 26(1), 54-64.

Montoya, P., Schandry, R., & Müller, A. (1993). Heartbeat evoked potentials (HEP): topography and influence of cardiac awareness and focus of attention. Electroencephalography and Clinical Neurophysiology, 88(2), 163-172.

Park, H. D., & Tallon-Baudry, C. (2014). The neural subjective frame: from bodily signals to perceptual consciousness. Philosophical Transactions of the Royal Society B: Biological Sciences, 369(1641), 20130208.

Couto, B., Adolfi, F., Sedeño, L., Salles, A., Canales-Johnson, A., Alvarez-Abut, P., ... & Ibanez, A. (2015). Disentangling interoception: insights from focal strokes affecting the perception of external and internal milieus. Frontiers in Psychology, 6, 503.